Synthesis of a Core Trisaccharide as a Versatile Building Block for N-Glycans and Glycoconjugates

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Dedicated to Professor Lutz Tietze on the occasion of his 60th birthday

Abstract: N-Linked oligosaccharides from glycoproteins (N-glycans) can be conveniently assembled with a building block approach. A protected form of the core trisaccharide (β -mannosyl chitobiose) was identified as a key building block. The chitobiose part of the core trisaccharide was built from a glycosyl fluoride, which served as a precursor for the reducing GlcNAc azide and the inner GlcNAc moiety. β -Mannosylation

Keywords: β -mannosides · carbohydrates · glycosylation · glycoproteins · oligosaccharide was accomplished at the trisaccharide stage by intramolecular inversion of a β glucosyl chitobiose. The benzylidene protection of the β -mannoside and the azido group at the reducing end of the core trisaccharide allow modular synthesis of N-glycans and their glycoconjugates.

Introduction

Asparagine-linked oligosaccharides (N-glycans) are a common feature of eucaryotic proteins that are surface bound or secreted.^[1] The biological properties of glycoproteins are influenced by the structure of their oligosaccharides. Despite a wide knowledge of structural details of N-glycans little is known about detailed structure–activity relationships of entire glycoproteins.^[2] One of the few glycoproteins studied thoroughly is recombinant human erythropoietin (EPO), used in the treatment of cancer and kidney patients. The serum halflife of EPO and its modification (NESP) can be tuned by the extent of N-glycosylation.^[3]

A combinatorial biochemical machinery is installing N-glycans as a posttranslational modification leading to a large variety of possible oligosaccharide structures. The resulting glycoproteins are therefore heterogeneous mixtures of individual glycoforms. The diversity of natural N-glycans is thus impairing the isolation of these compounds from glycoproteins.^[4-6] Therefore, the synthesis of N-glycans has been used to overcome the shortage of material for biological studies. While only few groups pioneered the chemical synthesis of

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Gebäude NW1, 95440 Bayreuth (Germany) Fax: (+49)921-55-5365 E-mail: Carlo.Unverzagt@uni-bayreuth.de N-glycans,^[7-9] the last decade has seen considerable increase in activity.^[10–15] Up to now several strategies have been developed leading to complete N-glycans or partial structures.^[16] The key challenges of the syntheses have been the construction of the β -mannoside^[17] followed by the incorporation of terminal sialic acid and the deprotection of the final products. Besides the chemical approaches the core trisaccharide has also been synthesized by enzymatic methods.^[18]

Results and Discussion

The total synthesis of N-glycans and their derivatives can be facilitated by combining chemical and enzymatic approaches.^[19] Our initial goal in N-glycan synthesis was the heptasaccharide – asparagine conjugate $\mathbf{1}$,^[20] which can serve both as a building block for glycopeptides^[21] and as an acceptor for enzymatic elongation of the sugar chains.^[19] Retrosynthetic analysis of 1 (Scheme 1) suggested a division into building blocks of general applicability.^[22] A protected core trisaccharide was desired which would permit good coupling to various building blocks for the antennae and allow the convenient construction of glycoconjugates through an amino function at the reducing end. Furthermore, the protected core trisaccharide should be available in amounts over 10 g on a typical laboratory scale to yield sufficient quantities of N-glycans after deprotection. The building block meeting these requirements was found in trisaccharide 2 (see Scheme 1). An optimized approach to this versatile compound is described in the following.

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- 1369



Scheme 1. Retrosynthetic analysis of the heptasaccharide-asparagine 1 suggests the disconnection to the core trisaccharide 2 as the key building block for N-glycans.

Synthesis of chitobiosylazide: The early introduction of an azido functionality^[23, 24] at the reducing end of the core trisaccharide **2** is advantageous because this moiety remains neutral under most reaction conditions applied in carbohydrate chemistry. Glycosyl azides can also be converted into glycosyl donors after derivatization.^[25, 26] The stable azido group at the anomeric center protects the labile amino function required for the attachment of aspartic acid^[27] or spacers. Glycosyl azides are obtained from the corresponding fluorides^[28] in high yield. The latter can be generated conveniently from thioglycosides.^[29, 30]

The starting material for the intermediate chitobiosyl azide **13** (Scheme 2) was the thioglycoside **3**,^[31, 32] a useful intermediate, which is accessible on a 100 g scale in a five-step sequence starting from glucosamine hydrochloride. First, the hydroxyl group was acetylated and the resulting thioglycoside **4** was converted to fluoride **6** with *N*-bromosuccinimide (NBS) and HF/pyridine according to Nicolaou.^[29] When activated with BF₃·Et₂O the fluoride^[28] **6** readily reacted with trimethylsilylazide to give the β -azide **8**. The deacetylation using potassium methoxide in dioxane/methanol 1:1 proceeded slowly and gave only 67 % yield. A nearly identical

 $R_{\rm f}$ value of compounds 8 and 10 required monitoring of the reaction by ¹H NMR spectroscopy. The cleavage appears to be sterically hindered^[33] and led to phthalimido ring opening during the prolonged reaction time. Similar difficulties occurred for the deacetylations at the di- and trisaccharide stage, which suggested the synthesis of an alternative set of building blocks carrying chloroacetyl groups. After introduction of the chloroacetyl moiety^[34] (5) the following transformations to fluoride 7 and azide 9 pro-

ceeded analogously in yields over 90%. With a base lability increased by three orders of magnitude,^[35] a dechloroacetylation occurs much faster than the cleavage of the corresponding acetate. The improved cleavage conditions permitted the deprotection of compound **9** to furnish the acceptor **10** in 92% yield.

Acceptor 10 was treated with the glycosylfluorides 6 and 7 activated by BF₃•Et₂O to give the chitobiosyl azides 11 and 12 in good yields. In the following deacylation step the chloroacetyl moiety could be readily removed. Compound 13 is equipped with a combination of protective groups suitable for the construction of the β -mannoside. The neighboring benzyl groups increase the reactivity^[36] of the OH-4" group; the phthalimido groups are known to be inert during nucleophilic substitutions and glycosylation reactions.

Synthesis of β **-mannosides**: The β -mannosidic linkage^[17] was established following a procedure introduced by Kunz.^[32, 37, 38] This method is based on the intramolecular inversion of 4,6-benzylidenated β -glucosides, which excludes unwanted anomeric or epimeric side products. For the synthesis of N-glycans with title compound **2** it was highly advantageous



that the β -mannosyl moiety was not blocked by permanent protective groups, for example benzyl ethers, but could be easily debenzylidenated at the desired time under mild conditions.^[32]

To introduce the β -glucosyl moiety the Kunz group used a glucosyl bromide,^[32] which gave also orthoesters under glycosylation conditions. Therefore, we converted peracetylated 3-phenylcarbamoyl glucose **14**^[32] to the anomeric fluoride **15** by reaction with HF/pyridine (Scheme 3).^[39] Elongation of the chitobiosyl acceptor **13** with the donor **15** under BF₃·Et₂O activation yielded 85% of the

 $\begin{array}{l} \label{eq:scheme 2. a) Ac_2O/pyridine (quant.); b) chloroacetic anhydride/pyridine, CH_2Cl_2 (93 \%); c) HF/pyridine, NBS, CH_2Cl_2 (93 \% 6), (96 \% 7); d) TMS-N_3, BF_3 \bullet OEt_2, CH_2Cl_2 (91 \% 8), (92 \% 9); e) MeOH, dioxane, NaOMe (67 \%); f) K_2CO_3, MeOH, CH_2Cl_2 (92 \%); g) BF_3 \bullet OEt_2, CH_2Cl_2 (83 \%); h) BF_3 \bullet OEt_2, CH_2Cl_2 (86 \%); i) K_2CO_3, MeOH, dioxane (79 \%); k) K_2CO_3, MeOH, CH_2Cl_2 (94 \%). \end{array}$

1370 —

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Scheme 3. a) HF/pyridine (56%); b) **13**, BF₃·OEt₂, CH₂Cl₂ (85%); c) 1) K₂CO₃, MeOH, dioxane, 2) benzaldehyde dimethylacetal, *p*-TosOH, CH₃CN (44% over two steps).

 β -gluco configured trisaccharide **16**. Prior to the inversion of the β -glucoside to the β -mannoside the 2-OH" group had to be selectively activated. Removal of the acetates and regio-selective benzylidenation of the intermediate triol gave the trisaccharide **17**. This compound was suitably functionalized for the ensuing inversion sequence to the β -mannoside.

Unfortunately, compound **17** was obtained in only 44% yield after the two-step reaction. The main reason was again a difficult removal of acetyl groups, in particular of the sterically hindered 2"-acetate.^[40] It is known that the deprotection of esters in the vicinity of sterically demanding residues is impaired. During the prolonged reaction times (1.5 d) the base labile phthalimido groups were also cleaved.

To circumvent these difficulties the novel β -glucosyl donor **20** (Scheme 4) was constructed, which was already equipped with a benzylidene acetal and carried an easily removable chloroacetyl group^[41] in position 2. Thus, the critical change of protective groups at the trisaccharide stage was avoided. In contrast to 3-phenylcarbamoyl β -glucosides, the free 3-phe-

nylcarbamoyl glucose **18** could not be acetalized selectively with benzaldehyde dimethylacetal. However, the traditional procedure with benzaldehyde in the presence of zinc chloride avoided the overreaction and gave the crystalline benzylidene acetal **19**. After chloroacetylation the anomeric center was liberated chemoselectively with hydrazine acetate and converted to the trichloroacetimidate **20** according to Schmidt.^[42] The imidate **20** could be converted to the β -fluoride **21**, which was formed stereoselectively through neighboring-group participation.

Initial experiments to glycosylate the chitobiosyl azide 13 with the imidate 20 showed only little conversion when BF₃. Et₂O was employed. Use of trimethylsilyl triflate^[42] (TMSOTf) gave the β -glucoside 22 in 62 % yield. The quality of the reagent was crucial for the success of the reaction. Only freshly opened trimethylsilyl triflate could suppress orthoester formation, which suggests that the deactivating 2-chloroacetyl moiety allows for the reaction to proceed via an intermediate orthoester. Furthermore, the yields were lowered by trimethylsilylation of the acceptor 13, which was desilylated and recycled after large-scale reactions. These problems were avoided using the fluoride 21 as a donor. The glucosyl fluoride could be activated with BF3 • Et2O and did not show formation of the orthoester. From compounds 21 and 13 the chloroacetylated trisaccharide 22 was obtained in 45% yield, which was conveniently deacylated (17) by mild base treatment. For the removal of the chloroacetyl moiety alternative methods^[43] are known, however, the rapid and easily adjustable procedure favored the use of potassium carbonate in methanol/dichloromethane.

The conversion of the β -gluco configured trisaccharide **17** to the β -mannoside **2** was achieved by a four-step procedure. This reaction sequence was previously established for monoor disaccharides^[32] and was successfully applied to the trisaccharide **17**. First, the 2"-OH function was converted to a good leaving group with trifluoromethanesulfonic acid anhydride and pyridine (Scheme 5). The intermediate triflate **17a** was substituted by an intramolecular nucleophilic attack of the neighboring carbonyl group of the phenylcarbamoyl moiety. This was achieved by warming a solution of the triflate in DMF/pyridine to 65 °C. The initially formed β -manno configured iminocarbonate **17b** was unstable and was con-



Scheme 4. a) Benzaldehyde, $ZnCl_2$ (34%); b) 1) chloroacetic anhydride/pyridine, CH_2Cl_2 , 2) hydrazine acetate, DMF; 3) trichloroacetonitrile, DBU, CH_2Cl_2 (over three steps 55%); c) HF/pyridine, CH_2Cl_2 (91%); d) TMSOTf, CH_2Cl_2 (62%); e) $BF_3 \cdot OEt_2$, CH_2Cl_2 (45%); f) K_2CO_3 , $MeOH/CH_2Cl_2$ (89%).

- 1371



Scheme 5. 1) Tf₂O, pyridine, CH_2Cl_2 ; 2) DMF, pyridine, 65 °C; 3) AcOH, dioxane, water; 4) NaOMe, MeOH, CH_2Cl_2 (overall yield 70%).

verted to the carbonate **17 c** by mild acid hydrolysis. Deprotection of the carbonate **17 c** by methanolysis gave the desired core trisaccharide **2** in 70% yield over the reaction sequence.^[22] The inversion at C-2" was conducted as a one-pot reaction and every step was monitored by TLC. NMR spectroscopy confirmed the newly formed β -mannosidic linkage via the C-1"/H-1" coupling constant of 163.3 Hz^[44, 45] determined from an HMQC spectrum without decoupling.^[46]

Following this approach the protected core trisaccharide **2** was obtained in overall amounts of more than 10 g. This building block has proven to be a key compound in the synthesis of several families of natural N-glycans^[15, 47] with different substitution patterns.

Experimental Section

General methods: Solvents were dried according to standard methods. Melting points were determined on a Büchi apparatus and are uncorrected. Optical rotations were measured on a Perkin–Elmer 241 polarimeter at 589 nm. NMR spectra were recorded on Bruker AC 250 and AMX 500 instruments. Coupling constants are reported in Hz. For mass spectra a Varian CH5 instrument was used in the fast atom bombardment mode (FAB) with a thioglycerine/HOAc (MB) or a *m*-nitrobenzylalcohol matrix (NBA). ESI-TOF mass spectra were recorded on a Micromass LCT instrument coupled to an Agilent 1100 HPLC. Flash chromatography was performed on silica gel 60 (230–400 mesh, Merck Darmstadt). The reactions were monitored by thin-layer chromatography on coated aluminum plates (silica gel 60 GF₂₅₄, Merck Darmstadt). Spots were detected by UV light or by charring with a 1:1 mixture of $2 \times H_2SO_4$ and 0.2% resorcine monomethyl ether in ethanol.

Ethyl 4-O-acetyl-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (4): Compound 3^[31, 32] (13.31 g, 24.9 mmol) was dissolved in pyridine (20 mL) and acetic anhydride (10 mL). After 20 h at ambient temperature the reaction mixture was concentrated and codistilled three times with toluene (50 mL). The residue was dried in high vacuo to yield 4 (14.35 g, quant.). $[\alpha]_D^{23} = +77.5^\circ$ (c = 1.0 in CH₂Cl₂); $R_f = 0.34$ (hexane/ acetone 2:1); ¹H NMR (250 MHz, [D₆]DMSO): δ = 7.87 - 7.79 (m, 4 H, Pht), 7.25 (m, 5H, Ph), 6.92 (m, 5H, Ph), 5.30 (d, $J_{12} = 10.4$ Hz, 1H, H-1 β), 5.00 (dd, $J_{4,5} = 9.8$ Hz, 1H, H-4), 4.56–4.44 (m, 3H, CH₂O), 4.35 (dd, $J_{3,4} =$ 8.1 Hz, 1H, H-3), 4.26 (d, $J_{gem} = 12.2$ Hz, 1H, CH₂O), 4.05 (dd, $J_{2,3} =$ 10.4 Hz, 1 H, H-2), 3.90-3.83 (m, 1 H, H-5), 3.59-3.47 (m, 2 H, H-6a,b), 2.57 (m, 2H, SCH₂), 2.02 (s, 3H, OAc), 1.09 (t, J = 7.4 Hz, CH₃); ¹³C NMR (62.5 MHz, $[D_6]DMSO$): $\delta = 168.8$ (C=O Ac), 166.9, 166.5 (C=O, Pht), 137.6, 136.9 (C-i Ph), 134.3 (C-4/5 Pht), 130.1 (C-1/2 Pht), 127.6, 127.4, 127.0, 126.8 (C-Ar), 122.9 (C-3/6 Pht), 80.0 (C-1), 77.1 (C-3), 76.1 (C-5), 73.0, 71.08 (CH₂O), 70.9 (C-4), 68.2 (C-6), 53.9 (C-2), 23.4 (CH₂), 20.7 (OAc), 15.0 (CH₃); elemental analysis calcd (%) for C₃₂H₃₃NO₇S (575.68): C 66.77, H 5.78, N 2.43; found: C 66.69, H 5.56, N 2.57.

fluoride (6): Thioglycoside 4 (12.6 g, 21.9 mmol) was dissolved in dichloromethane (130 mL) and cooled to 0°C. N-Bromosuccinimide (5.83 g, 32.7 mmol) was added to the stirred solution followed by HF/pyridine complex (12.6 mL). The reaction was complete after 10 min (TLC: hexane/ acetone 1.5:1). Subsequently, dichloromethane (130 mL) was added and the reaction mixture was extracted in a Teflon separatory funnel with water, aq HCl $(3 \times)$ and aq Na₂CO₃. The organic phase was dried (MgSO₄), concentrated and purified by flash chromatography (hexane/acetone 2:1) to yield 6 (10.83 g, 92.7%). The product was recrystallized from acetone/ hexane. M.p. 100 °C; $R_{\rm f} = 0.45$ (hexane/acetone 1.5:1); $[\alpha]_{\rm D}^{23} = +76.4^{\circ}$ (c = 0.5 in CH₂Cl₂); ¹H NMR (250 MHz, $[D_6]DMSO$): $\delta = 7.88$ (m, 4H, Pht), 7.35 (m, 5H, Ph), 6.93 (m, 5H, Ph), 5.93 (dd, $J_{12} = 8.0$, $J_{1F} = 53.6$ Hz, 1H, H-1*β*), 5.10 (dd, *J*_{4.5} = 9.5 Hz, 1 H, H-4), 4.57-4.44 (m, 3 H, CH₂O), 4.39 (dd, $J_{2,3} = J_{3,4} = 9.9$ Hz, 1 H, H-3), 4.27 (d, $J_{gem} = 12.0$ Hz, 1 H, CH₂O), 4.14 (m, 1H, H-2), 4.04 (m, 1H, H-5), 3.58 (m, 2H, H-6a,b), 2.05 (s, 3H, OAc); ¹³C NMR (62.5 MHz, $[D_6]$ DMSO): $\delta = 169.2, 167.2$ (C=O), 137.9, 137.3 (C-*i* Ph), 134.9 (C-4/5 Pht), 130.5 (C-1/2 Pht), 128.2-127.5 (C-Ar), 123.5 (C-3/6 Pht), 104.0 (d, $J_{C-1,F} = 212.3$ Hz, C-1), 75.9 (d, $J_{C-3,F} = 9.7$ Hz, C-3), 73.4 (CH₂O), 72.5 (C-5), 72.4 (CH₂O), 70.7 (C-4), 68.1 (C-6), 55.1 (d, J_{C-2,F} = 21.8 Hz, C-2), 20.6 (OAc); elemental analysis calcd (%) for C₃₀H₂₈FNO₇ (533.55): C 67.53, H 5.29, N 2.63; found: C 67.62, H 5.48, N 2.52.

4-O-Acetyl-3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosylazide (8): Trimethylsilylazide (2.35 mL, 17.7 mmol) was added to a stirred solution of fluoride 6 (4 g, 75 mmol) in dry diablare methane (70 mL). The

solution of fluoride 6 (4 g, 7.5 mmol) in dry dichloromethane (70 mL). The reaction was initiated with BF3 · OEt2 (300 µL, 2.4 mmol). After complete reaction (TLC: hexane/acetone 1.5:1) dichloromethane was added followed by extraction with dilute K2CO3. The organic phase was dried (MgSO₄), evaporated and purified by flash chromatography (hexane/ acetone 2:1) to afford **8** (3.81 g, 91.3%). $[\alpha]_{D}^{23} = +49.5^{\circ}$ (c = 0.5 in dichloromethane); $R_f = 0.48$ (hexane/acetone 1.5:1); ¹H NMR (250 MHz, $[D_6]$ DMSO): $\delta = 7.9 - 7.7$ (m, 4H, Pht), 7.4 - 7.25 (m, 5H, Ph), 6.94 (m, 5H, Ph), 5.54 (d, $J_{1,2} = 9.5$ Hz, 1 H, H-1 β), 5.04 (dd, $J_{3,4} = J_{4,5} = 9.5$ Hz, 1 H, H-4), 4.57 - 4.47 (m, 3 H, CH₂O), 4.37 (dd, $J_{2,3} = 10.1$ Hz, 1 H, H-3), 4.26 (d, $J_{gem} = 10.1$ Hz, 1 Hz, 1 H, H-3), 4.26 (d, $J_{gem} = 10.1$ 12.0 Hz, 1 H, CH₂O), 3.98 (m, 1 H, H-5), 3.94 (dd, 1 H, H-2), 3.6-3.5 (m, 2 H, H-6a,b), 2.03 (s, 3 H, OAc); ¹³C NMR (125 MHz, $[D_6]DMSO$): $\delta =$ 169.3, 167.0 broad (C=O), 138.1, 137.4 (C-i Ph), 134.9 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.2, 127.9, 127.6, 127.50, 127.47, 127.42 (C-Ar), 123.5 (C-3/6 Pht), 85.0 (C-1), 76.4 (C-3), 74.6 (C-5), 73.5 (CH₂O), 71.1 (C-4), 72.4 (CH₂O), 71.1 (C-4), 68.3 (C-6), 54.6 (C-2), 20.7 (OAc); elemental analysis calcd (%) for C₃₀H₂₈N₄O₇ (556.57): C 64.74, H 5.07, N 10.07; found: C 64.71, H 5.00, N 10.00.

3,6-Di-O-benzyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosylazide (10): a) Starting from 8: NaOMe (100 mg) was added to a stirred solution of acetate 8 (3.7 g, 6.65 mmol) in dioxane/methanol 1:1 (40 mL). The course of the reaction was monitored by ¹H NMR spectroscopy following the disappearance of the signal of the acetyl group. Two further portions of sodium methylate (100 mg each) were added after 30 min intervals. After

complete reaction the solution was neutralized with the acidic ion exchange resin Amberlyst $15 \,\mathrm{H^+}$, filtered and concentrated. The residue was purified by flash chromatography (hexane/acetone 2.2:1) and gave **10** (2.3 g, 67.2 %).

b) Starting from 9: K₂CO₃ (200 mg) was added to a stirred solution of chloroacetate 9 (12.0 g, 20.3 mmol) in methanol/dichloromethane 1:2 (120 mL). After complete reaction (TLC: hexane/ethyl acetate 1.5:1) the solution was filtered, neutralized with acidic acid and concentrated. The residue was purified by flash chromatography (hexane/ethyl acetate 5:3) and gave 10 (9.6 g, 91.9%). $[\alpha]_D^{23} = +17.2^{\circ}$ (c = 0.5 in CH₂Cl₂); $R_f = 0.45$ (hexane/ethyl acetate 1.5:1); ¹H NMR (500 MHz, $[D_6]DMSO$): $\delta = 7.9 - 7.7$ (m, 4H, Pht), 7.4 - 7.33 (m, 5H, Ph), 6.93 - 6.86 (m, 5H, Ph), 5.74 (d, J_{4.0H} = -6.86 (m, 5H, Ph))7.2 Hz, 1 H, OH-4), 5.42 (d, $J_{1,2} = 9.5$ Hz, 1 H, H-1 β), 4.78 (d, $J_{gem} = 12.2$ Hz, 1 H, CH₂O), 5.58 (s, 2 H, CH₂O), 4.42 (d, J_{gem} = 12.2 Hz, 1 H, CH₂O), 4.14 $(dd, J_{3,4} = J_{2,3} = 9.6 Hz, 1 H, H-3), 3.86 - 3.82 (m, 2 H, H-6a, H-2), 3.75 - 3.68$ (m, 2 H, H-6b, H-5), 3.54 (ddd, $J_{4.5} = 8.5$ Hz, 1 H, H-4); ¹³C NMR (125 MHz, $[D_6]DMSO$: $\delta = 167.3$ (C=O), 138.4, 138.1 (C-*i* Ph), 134.7 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.2-127.1 (C-Ar), 123.5 (C-3/6 Pht), 84.9 (C-1), 78.3 (C-3), 77.8 (C-5), 73.6, 72.3 (CH₂O), 71.1 (C-4), 68.9 (C-6), 54.6 (C-2); elemental analysis calcd (%) for $C_{28}H_{26}N_4O_6\,(514.53)$: C 65.36, H 5.09, N 10.89; found: C 65.45, H 5.33, N 10.78.

Ethyl 3,6-di-O-benzyl-4-O-chloroacetyl-2-deoxy-2-phthalimido-1-thio-β-**D-glucopyranoside (5)**: Compound **3**^[31, 32] (97 g, 181.8 mmol) and pyridine (100 mL) were dissolved in dichloromethane (500 mL) and cooled to 0 °C. Chloroacetic acid anhydride (45 g, 263.2 mmol) was added and the reaction was maintained at 0 °C until the starting material was consumed (TLC: hexane/ethyl acetate 1:1). Subsequently, the solution was diluted with dichloromethane (500 mL) and extracted with dilute K₂CO₃, dilute HCl $(3 \times)$ and dilute K₂CO₃. The organic phase was dried (MgSO₄), concentrated and purified by flash chromatography (hexane/acetone 2:1) to give 5 (103.4 g, 93.2%). $[a]_{D}^{23} = +50.9^{\circ}$ (c=1 in dichloromethane); $R_{f} = 0.66$ (hexane/EtOAc 1:1); ¹H NMR (250 MHz, $[D_6]DMSO$): $\delta = 7.9 - 7.7$ (m, 4H, Pht), 7.34 (m, 5H, Ph), 6.92 (m, 5H, Ph), 5.30 (d, $J_{1,2} = 10.4$ Hz, 1H, H-1β), 5.10 (dd, J_{4.5} = 9.5 Hz, 1 H, H-4), 4.6-4.35 (m, 6 H, CH₂O, CH₂Cl, H-3), 4.26 (d, J_{gem} = 12.2 Hz, 1 H, CH₂O), 4.07 (dd, J_{2.3} = 10.3 Hz, 1 H, H-2), 3.92 (m, 1H, H-5), 3.58 (m, 2H, H-6a,b), 2.68-2.47 (m, 2H, SCH₂), 1.09 (t, J = 7.4 Hz, CH₃); ¹³C NMR (62.5 MHz, [D₆]DMSO): $\delta = 167.4$, 167.0, 166.4 (C=O), 138.4, 137.4 (C-i Ph), 134.9 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.2-127.4 (C-Ar), 123.5 (C-3/6 Pht), 80.5 (C-1), 77.5 (C-3), 76.3 (C-5), 73.7 (CH₂O), 74.1 (C-4), 72.5 (CH₂O), 68.5 (C-6), 54.4 (C-2), 41.0 (CH₂Cl), 23.5 (CH₂), 15.1 (CH₃); elemental analysis calcd (%) for C₃₂H₃₂ClNO₇S (610.12): C 63.0, H 5.29, N 2.30; found: C 62.85, H 5.48, N 2.07.

3,6-Di-O-benzyl-4-O-chloroacetyl-2-deoxy-2-phthalimido-β-D-glucopyranosylfluoride (7): Thioglycoside 5 (112.0 g, 183.6 mmol) was dissolved in dichloromethane (800 mL) and cooled to 0°C. N-Bromosuccinimide (42.5 g, 239.8 mmol) was added to the stirred solution followed by HF/ pyridine complex (20 mL). The reaction was complete after 20 min (TLC: hexane/acetone 1.5:1). Subsequently, the reaction mixture was transferred to a Teflon separatory funnel and extracted with a mixture of water and ice $(2 \times)$, dilute HCl $(3 \times)$ and aq Na₂CO₃. The organic phase was dried (MgSO₄), concentrated and purified by flash chromatography (hexane/ ethyl acetate 2:1) to yield 7 (99.6 g, 95.5 %). The product was recrystallized from acetone/hexane. M.p. 115 °C; $[\alpha]_{D}^{23} = +75.4^{\circ}$ (c = 1 in CH₂Cl₂); $R_{f} =$ 0.33 (hexane/acetone 2:1); ¹H NMR (250 MHz, $[D_6]DMSO$): $\delta = 7.85$ (m, 4H, Pht), 7.35 (m, 5H, Ph), 6.93 (m, 5H, Ph), 5.96 (dd, $J_{1,2} = 7.8$, $J_{1,F} = 7.8$ 53.4 Hz, 1 H, H-1 β), 5.22 (dd, $J_{3,4} = J_{4,5} = 9.4$ Hz, 1 H, H-4), 4.6 – 4.4 (m, 6 H, CH₂O, CH₂Cl, H-3), 4.27 (d, J_{gem} = 12.0 Hz, 1 H, CH₂O), 4.2 (m, 1 H, H-2), 4.13 (m, 1H, H-5), 3.64 (m, 2H, H-6a,b); ¹³C NMR (62.5 MHz, $[D_6]DMSO$): $\delta = 167.2$, 166.4 (C=O, Ac, Pht), 137.9, 137.2 (C-*i* Ph), 134.9 (C-4/5 Pht), 130.5 (C-1/2 Pht), 128.2, 127.9, 127.6, 127.5 (C-Ar), 123.6 (C-3/6 Pht), 104.5 (d, $J_{C-1F} = 212.4$ Hz, C-1), 75.3 (d, $J_{C-3F} = 9.7$ Hz, C-3), 73.6 (CH₂O), 72.5 (CH₂O), 72.3 (C-4), 72.2 (C-5), 68.0 (C-6), 55.1 (d, $J_{C-2,F} =$ 22.0 Hz, C-2), 40.9 (CH₂Cl); elemental analysis calcd (%) for C₃₀H₂₇ClFNO₇×H₂O (586.01): C 61.49, H 4.99, N 2.39; found: C 61.82, H 4.94, N 2.66.

 $\textbf{3,6-Di-}\textit{O-benzyl-4-}\textit{O-chloroacetyl-2-deoxy-2-phthalimido-}\beta\textbf{-}\textbf{D-glucopyra-}$

nosylazide (9): TMS-N₃ (20 mL, 150.6 mmol) followed by BF₃ · OEt₂ (2 mL, 16.3 mmol) was added to a stirred suspension of fluoride **7** (40.0 g, 70.4 mmol) and ground molecular sieves 4 Å (20 g) in dry dichloromethane (400 mL). After complete reaction (TLC: hexane/acetone 2:1) the mixture was diluted with dichloromethane and filtered. The organic phase was

washed with aq K₂CO₃, dried (MgSO₄) and concentrated. The residue was purified by flash chromatography (hexane/acetone 2:1) to yield **9** (38.2 g, 91.8%). Crystals were obtained by trituration of the product with methanol. M.p. 86 – 91 °C; $[\alpha]_{D^3}^{T^3} = +45.0^{\circ}$ (c = 0.5 in dichloromethane); $R_t = 0.37$ (hexane/acetone 2:1); ¹H NMR (250 MHz, [D₆]DMSO): $\delta = 7.85$ (m, 4H, Pht), 7.35 (m, 5H, Ph), 6.92 (m, 5H, Ph), 5.57 (d, $J_{1,2} = 9.4$ Hz, 1 H, H-1β), 5.15 (dd, $J_{4,5} = 9.5$ Hz, 1 H, H-4), 4.6 – 4.4 (m, 6H, CH₂O, CH₂Cl, H-3), 4.25 (d, $J_{gem} = 12.0$ Hz, 1 H, CH₂O), 4.03 (m, 1 H, H-5), 3.97 (dd, $J_{2,3} = 10.6$ Hz, 1 H, H-2), 3.63 (m, 2 H, H-6a,b); ¹³C NMR (62.5 MHz, [D₆]DMSO): $\delta = 167.2$, 166.4 (C=O, CIAc, Pht), 138.0, 137.3 (C-*i* Ph), 134.9 (C-4/5 Pht), 130.5 (C-1/2 Pht), 128.2, 127.9, 127.5 (C-Ar), 123.5 (C-3/6 Pht), 84.9 (C-1), 76.2 (C-3), 74.3 (C-5), 73.6 (CH₂O), 72.6 (C-4), 72.4 (CH₂O), 68.0 (C-6), 54.5 (C-2), 40.9 (CH₂Cl); ESI-MS: m/z: calcd for C₃₀H₂₇ClN₄O₇: 590.16; found: 613.26 [M+Na]⁺.

 $O-(4-O-Acetyl-3,6-di-O-benzyl-2-deoxy-2-phthalimido-\beta-D-glucopyrano-syl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl-$

azide (11): A suspension of azide 10 (3.0 g, 5.8 mmol), fluoride 6 (3.9 g, 7.3 mmol) and ground molecular sieves 4 Å (7 g) in dry dichloromethane (70 mL) was stirred for 30 min at room temperature. BF3 \cdot OEt2 (150 µL, 1.2 mmol) was added and the reaction was allowed to proceed for 1 h (TLC: hexane/acetone 1.5:1). The suspension was filtered over Celite, washed with dichloromethane followed by extraction with aq Na₂CO₃. The organic phase was dried (MgSO₄), concentrated and the residue was purified by flash chromatography (hexane/acetone 2.3:1) to give 11 (5.0 g, 83.4%). $[a]_{D}^{23} = +26.5^{\circ}$ (c = 0.5 in CH₂Cl₂); $R_{f} = 0.42$ (hexane/acetone 1.5:1); ¹H NMR (500 MHz, $[D_6]$ DMSO): $\delta = 8.0 - 7.7$ (m, 8 H, Pht), 7.4 - 7.2 (m, 10H, Ph), 6.95-6.80 (m, 10H, Ph), 5.31 (d, $J_{1,2} = 7.2$ Hz, 1H, H-1' β), 5.29 (d, $J_{1,2} = 9.0$ Hz, 1 H, H-1 β), 5.01 (dd, $J_{3,4} = J_{4,5} = 9.5$ Hz, 1 H, H-4'), 4.81 (d, J_{gem} = 12.3 Hz, 1 H, CH₂O), 4.54-4.36 (m, 7 H, CH₂O, H-3'), 4.26 (d, $J_{\text{gem}} = 12.0 \text{ Hz}, 1 \text{ H}, \text{ CH}_2\text{O}), 4.12 - 4.07 \text{ (m, 3H, H-2', H-3, H-4)}, 3.80 \text{ (dd,}$ $J_{2,3} = 10.6$ Hz, 1H, H-2), 3.62 - 3.54 (m, 3H, H-5, H-5', H-6a'), 3.49 (d, $J_{\text{gem}} = 10.9 \text{ Hz}, 1 \text{ H}, \text{ H-6a}), 3.41 \text{ (dd}, J_{\text{vic}} = 5.5, J_{\text{gem}} = 11.2 \text{ Hz}, 1 \text{ H}, \text{ H-6b'}), 3.37 \text{ (dd}, J_{\text{vic}} = 3.8 \text{ Hz}, 1 \text{ H}, \text{ H-6b}), 2.03 \text{ (OAc)}; {}^{13}\text{C} \text{ NMR} \text{ (125 MHz}, 1 \text{ Hz})$ $[D_6]DMSO$): $\delta = 169.4$, 167.1 (C=O), 138.2, 138.0, 137.9, 137.4 (C-*i* Ph), 134.9, 134.8 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.2 - 127.0 (C-Ar), 123.5 (C-3/6 Pht), 96.4 (C-1'), 84.8 (C-1), 76.3 (C-3'), 76.1 (C-3), 75.6 (C-5), 75.1 (C-4), 73.7, 73.5 (CH₂O), 73.0 (C-5'), 72.5, 71.7 (CH₂O), 71.5 (C-4'), 68.3 (C-6'), 67.6 (C-6), 55.7 (C-2'), 54.6 (C-2), 20.6 (OAc); elemental analysis calcd (%) for C58H53N5O13 × 0.5 H2O (1028.08): C 67.17, H 5.25, N 6.75; found: C 67.07, H 5.07. N 6.42.

O-(3,6-Di-O-benzyl-4-O-chloroacetyl-2-deoxy-2-phthalimido- β -D-gluco-pyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyra-

nosylazide (12): A suspension of azide 10 (13.0 g, 25.3 mmol), fluoride 7 (17.0 g, 29.9 mmol) and ground molecular sieves 4 Å (22 g) in dry dichloromethane (350 mL) was stirred for 30 min at room temperature. BF3 • OEt2 (1.46 mL, 11.9 mmol) was added and the reaction was allowed to proceed for 1 h (TLC: hexane/acetone 1.5:1). The suspension was filtered over Celite, washed with dichloromethane followed by extraction with aq Na₂CO₃. The organic phase was dried (MgSO₄), concentrated and the residue was purified by flash chromatography (hexane/acetone 2:1) furnishing **12** (23.12 g, 86.1 %). $[\alpha]_{D}^{23} = +17.6^{\circ} (c = 0.5 \text{ in dichloromethane});$ $R_{\rm f} = 0.36$ (hexane/acetone 1.5:1); ¹H NMR (500 MHz, [D₆]DMSO): $\delta =$ 7.9-7.7 (m, 8H, Pht), 7.4-7.2 (m, 10H, Ph), 6.92-6.79 (m, 10H, Ph), 5.31 $(d, J_{1,2} = 7.9 \text{ Hz}, 1 \text{ H}, \text{H}-1'\beta), 5.29 (d, J_{1,2} = 9.2 \text{ Hz}, 1 \text{ H}, \text{H}-1\beta), 5.12 (dd, J_{3,4} = 0.2 \text{ Hz})$ 12.0 Hz, 1 H, CH₂O), 4.50 (d, $J_{gem} = 12.0$ Hz, 1 H, CH₂O), 4.44–4.38 (m, 7 H, CH₂O, CH₂Cl, H-3'), 4.26 (d, J_{gem} = 12.1 Hz, 1 H, CH₂O), 4.16 – 4.05 (m, 3 H, H-2', H-3, H-4), 3.81 (dd, *J*_{2,3} = 10.5 Hz, 1 H, H-2), 3.63 – 3.55 (m, 3 H, H-5, H-5', H-6a'), 3.49 (d, $J_{gem} = 10.8$ Hz, 1 H, H-6a), 3.45 (dd, $J_{vic} = 5.0$, $J_{\text{gem}} = 11.1 \text{ Hz}, 1 \text{ H}, \text{ H-6b'}), 3.37 \text{ (dd, } J_{\text{vic}} = 3.7 \text{ Hz}, 1 \text{ H}, \text{ H-6b}); {}^{13}\text{C} \text{ NMR}$ (125 MHz, [D₆]DMSO): δ = 168.1, 167.1, 166.5 (C=O), 138.3, 138.1, 138.0, 137.4 (C-i Ph), 134.9, 134.8 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.3-127.0 (C-Ar), 123.6, 123.4 (C-3/6 Pht), 96.5 (C-1'), 84.8 (C-1), 76.2 (C-3', C-3'), 75.6 (C-5), 75.2 (C-4), 73.8, 73.7 (CH₂O), 73.1 (C-4'), 72.7 (C-5'), 72.6, 71.7 (CH2O), 68.1 (C-6'), 67.6 (C-6), 55.8 (C-2'), 54.6 (C-2), 41.0 (CH2CI); elemental analysis calcd (%) for C58H52CIN5O13 (1062.53): C 65.56, H 4.93, N 6.59; found: C 65.39, H 4.89, N 6.50.

- 1373

potassium carbonate (350 mg). The suspension was filtered, neutralized with acetic acid and concentrated. The residue was purified by flash chromatography (hexane/acetone 2:1) and gave title compound **13** (3.1 g, 78.7 %).

b) Starting from 12: Disaccharide 12 (14.0 g, 13.2 mmol) was dissolved in methanol/dichloromethane 1:2 (120 mL) and stirred with potassium carbonate (30 mg). After complete reaction (TLC: hexane/acetone 1.5:1) the suspension was filtered, neutralized with acetic acid and concentrated. The residue was purified by flash chromatography (hexane/acetone 1.5:1) and yielded **13** (12.2 g, 93.7 %). $[\alpha]_{\rm D}^{23} = -3.3^{\circ}$ (c = 0.5 in CH₂Cl₂); $R_{\rm f} = 0.22$ (hexane/acetone 1.5:1); ¹H NMR (500 MHz, $[D_6]DMSO$): $\delta = 7.95 - 7.7$ (m, 8H, Pht), 7.4–7.2 (m, 10H, Ph), 6.95–6.80 (m, 10H, Ph), 5.63 (d, $J_{4,OH} =$ 7.2 Hz, 1 H, OH-4), 5.29 (d, $J_{1,2} = 9.1$ Hz, 1 H, H-1 β), 5.27 (d, $J_{1,2} = 7.1$ Hz, 1 H, H-1' β), 4.82 (d, J_{gem} = 12.2 Hz, 1 H, CH₂O), 4.77 (d, J_{gem} = 12.2 Hz, 1 H, CH₂O), 4.60 (d, $J_{gem} = 12.4$ Hz, 1H, CH₂O), 4.48 (d, $J_{gem} = 12.4$ Hz, 1H, CH₂O), 4.45-4.38 (m, 4H, CH₂O), 4.15 (dd, J_{3,4} = 8.4 Hz, 1H, H-3'), 3.98 $(dd, J_{23} = 10.5 Hz, 1 H, H-2'), 3.85 - 3.80 (m, 2 H, H-2, H-6a'), 3.60 (m, 1 H, H-2)$ H-5), 3.56-3.48 (m, 3H, H-6b', H-4', H-6a), 3.43 (m, 1H, H-5'), 3.37 (dd, $J_{\rm vic} = 3.6, J_{\rm gem} = 11.3$ Hz, 1 H, H-6b); ¹³C NMR (125 MHz, [D₆]DMSO): $\delta =$ 168.2, 167.1 (C=O), 138.8, 138.3, 138.1, 138.0 (C-i Ph), 134.8 (C-4/5 Pht), 130.8, 130.6 (C-1/2 Pht), 128.2-127.0 (C-Ar), 123.4 (C-3/6 Pht), 96.5 (C-1'), 84.9 (C-1), 78.4 (C-3'), 76.1 (C-3, C-5'), 75.8 (C-5), 74.8 (C-4), 73.6, 72.4, 71.71 (CH₂O), 71.65 (C-4'), 68.9 (C-6'), 67.5 (C-6), 55.9 (C-2'), 54.6 (C-2); elemental analysis calcd (%) for C56H51N5O12 (986.05): C 68.21, H 5.21, N 7.10: found: C 68.09. H 5.15. N 7.02.

$2,4,6\text{-}Tri\text{-}O\text{-}acetyl\text{-}3\text{-}O\text{-}(N\text{-}phenylcarbamoyl)\text{-}\alpha\text{-}D\text{-}glucopyranosylfluoride}$

(15): Acetate $14^{[32]}$ (11.0 g, 23.5 mmol) was dissolved in HF/pyridine (20 mL). After 20 h at ambient temperature the mixture was added to dichloromethane (200 mL) and ice (200 mL) in a Teflon separatory funnel. The organic phase was extracted with aq HCl $(3 \times)$, aq K₂CO₃ and dried (MgSO₄). After removal of the solvent the residue was purified by flash chromatography (hexane/acetone 2.5:1) and furnished fluoride 15 (5.6 g, 55.7%). $[\alpha]_{D}^{23} = +57.6^{\circ}$ (c = 1 in dichloromethane); $R_{f} = 0.38$ (hexane/ acetone 1.5:1); ¹H NMR (250 MHz, $[D_6]$ DMSO): $\delta = 9.8$ (s, 1 H, NH), 7.5 – 7.0 (m, 5 H, Ph), 5.94 (dd, $J_{1,F} = 52.9$, $J_{1,2} = 2.5$ Hz, 1 H, H-1 α), 5.32 (dd, $J_{2,3} = 2.5$ Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H-1 α), 5.32 (dd, J_{2,3} = 2.5 Hz, 1 H, H, $J_{3,4} = 9.9$ Hz, 1H, H-3), 5.15 (dd, $J_{4,5} = 8.8$ Hz, 1H, H-4), 5.06 (ddd, $J_{2,F} =$ 24.8 Hz, 1 H, H-2), 4.28 - 4.2 (m, 2 H, H-5, H-6a), 4.10 (dd, $J_{gem} = 14.4$, $J_{vic} = 14.4$ 4.5 Hz, 1H, H-6b), 2.04, 1.98 (2s, 9H, OAc); ¹³C NMR (125 MHz, $[D_6]DMSO$): $\delta = 169.9$, 169.4, 169.1 (C=O), 152.3 (C=O urethane), 138.6 (C-*i* Ph), 128.7, 122.8, 118.4 (C-Ar), 104.0 (d, J_{C-1,F} = 226.4 Hz, C-1), 69.7 (C-5), 69.4 (d, $J_{C2,F} = 24.2$ Hz, C-2), 69.3 (C-3), 67.2 (C-4), 61.4 (C-6), 20.4, 20.2 (OAc); elemental analysis calcd (%) for $C_{19}H_{22}FNO_9$ (427.38): C 53.4, H 5.19, N 3.28; found: C 53.51, H 5.10, N 3.44.

$O\-(2,4,6\-Tri-O\-acetyl-3\-O\-(N\-phenylcarbamoyl)\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl\-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl\-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl\-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl\-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl\-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-O\-benzyl\-2\-deoxy\-2\-phthalimido\-\beta\-D\-glucopyranosyl)\-(1 \rightarrow 4)\-O\-(3,6\-di\-D\-benzyl\-2\-deoxy\-2\-benzyl\-2\-deoxy\-2\-benzyl\-2\-deoxy\-2\-benzyl\-2$

 $(1 \rightarrow 4)$ -3,6-di-*O*-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosylazide (16): Disaccharide 13 (2.9 g, 2.9 mmol), fluoride 15 (1.95 g, 4.6 mmol) and ground molecular sieves 4 Å (3.4 g) in dry dichloromethane (50 mL) were stirred for 30 min at room temperature. BF3 • OEt2 (500 µL, 4.1 mmol) was added and the reaction was allowed to proceed for 7 h (TLC: hexane/ acetone 1.5:1). The suspension was filtered over Celite, washed with dichloromethane followed by extraction with aq Na2CO3. The organic phase was dried (MgSO₄), concentrated and the residue was purified by flash chromatography (hexane/acetone 2:1) to yield trisaccharide 16 (3.5 g, 85.4%). $[\alpha]_{D}^{23} = -7.5^{\circ}$ (c = 0.5 in dichloromethane); $R_{f} = 0.28$ (hexane/ acetone 1.5:1); ¹H NMR (500 MHz, $[D_6]DMSO$): $\delta = 9.7$ (s, 1 H, NH), 7.9-7.65 (m, 8H, Pht), 7.50-7.20 (m, 20H, Ar), 7.02-6.8 (m, 10H, Ar), 5.28 (d, $J_{1,2} = 9.5$ Hz, 1H, H-1 β), 5.27 (m, 1H, H-3"), 5.25 (d, $J_{1,2} = 8.5$ Hz, 1H, H-1' β), 4.95 (dd, $J_{3,4} = J_{4,5} = 9.6$ Hz, 1H, H-4"), 4.90-4.86 (m, 2H, H-1", H-2"), 4.83 – 4.78 (m, 2H, CH₂O), 4.63 (d, $J_{gem} = 12.3$ Hz, 1H, CH₂O), 4.57 $(d, J_{gem} = 12.3 \text{ Hz}, 1 \text{ H}, \text{CH}_2\text{O}), 4.41 \text{ (m}, 3 \text{ H}, \text{CH}_2\text{O}), 4.31 \text{ (d}, J_{gem} = 12.3 \text{ Hz},$ 1H, CH₂O), 4.20-4.06 (m, 4H, H-3', H-3, H-4, H-6a"), 4.03 (dd, J_{2,3}= 10.5 Hz, 1 H, H-2'), 3.94-3.89 (m, H-4', H-5", H-6b"), 3.83-3.78 (m, 2 H, H-2, H-6a'), 3.61-3.58 (m, 2H, H-5, H-6b'), 3.50 (dd, $J_{vic} < 1.0$, $J_{eem} =$ 11.6 Hz, 1H, H-6a), 3.4-3.3 (m, 2H, H-5, H-6b); ¹³C NMR (125 MHz, $[D_6]DMSO$): $\delta = 169.8$, 169.2, 168.9, 168.1, 167.1 (C=O), 152.42 (C=O) urethane), 138.7, 138.31, 138.25, 138.1, 138.0 (C-i Ar), 134.8 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.8, 128.3, 128.2, 127.8, 127.7, 127.5, 127.4, 127.2, 127.0 (C-Ar), 123.5 (C-3/6 Pht), 122.8, 118.5 (C-Ar), 99.8 (C-1"), 96.3 (C-1"), 84.9 (C-1), 78.4 (C-4'), 76.8 (C-3'), 76.1 (C-3), 75.6 (C-5), 75.0 (C-4), 74.7 (C-5'), 73.8, 73.7 (CH₂O), 72.4 (CH₂O, C-3"), 71.7 (CH₂O), 71.5 (C-2"), 70.6 (C-5"), 68.4 (C-4"), 67.6 (C-6), 67.5 (C-6'), 61.7 (C-6"), 55.8 (C-2'), 54.7 (C-2), 20.4, 20.3 (OAc); elemental analysis calcd (%) for $C_{75}H_{72}N_6O_{21}$ (1393.42): C 64.65, H 5.21, N 6.03; found: C 64.81, H 5.24, N 5.66.

4,6-O-Benzylidene-3-O-(N-phenylcarbamoyl)-D-glucopyranoside (19): Compound 18^[32] (101 g, 337 mmol) and anhydrous zinc chloride (25 g) were suspended in benzaldehyde (270 mL) and stirred for 3 d at ambient temperature (TLC: hexane/acetone 1:1). The reaction was added to ethyl acetate/water 1:1 (1.6 L). After thorough extraction a first fraction of product crystallized. The crystals were filtered off and washed with water and diethyl ether. The organic phase was dried (MgSO4) and diluted with hexane. A second crop of crystalline product was filtered off and washed with water and diethyl ether. The fractions of crystalline 19 were combined (45 g, 34.4 %). M.p. 179 °C; $[\alpha]_{D}^{23} = +2.2^{\circ}$ (1, MeOH/DMF 5:1); $R_{f} = 0.42$ (hexane/acetone 1:1); in DMSO compound 19 was present as a mixture of anomers ($\alpha:\beta=2:1$); α -anomer: ¹H NMR (500 MHz, [D₆]DMSO): $\delta=9.61$ (s, 1 H, NH), 7.5 – 7.2 (m, 9 H, Ph), 6.95 (m, 1 H, Ph), 6.88 (d, J_{1,OH} = 4.8 Hz, 1H, OH-1), 5.59 (s, 1H, =CH-Ph), 5.15 (dd, J_{2,3}=J_{3,4}=9.7 Hz, 1H, H-3), 5.08 (dd, $J_{1,2} = 3.6$ Hz, 1 H, H-1 α), 5.02 (d, $J_{2,OH} = 7.7$ Hz, 1 H, OH-2), 4.16 $(dd, J_{gem} = 10.2, J_{vic} = 4.5 Hz, 1 H, H-6a), 3.93 (m, 1 H, H-5), 3.74 (dd, J_{vic} = 10.2 Hz)$ 10.2 Hz, 1 H, H-6b), 3.62 (m, 1 H, H-4), 3.55 (m, 1 H, H-2); β-anomer: ¹H NMR (500 MHz, [D₆]DMSO): $\delta = 9.63$ (s, 1 H, NH), 7.5 – 7.2 (m, 9 H, Ph), 7.07 (d, J_{1.0H} = 6.5 Hz, 1 H, OH-1), 6.95 (m, 1 H, Ph), 5.60 (s, 1 H, =CH-Ph), 5.48 (d, $J_{2,OH} = 5.7$ Hz, 1H, OH-2), 4.99 (dd, $J_{2,3} = J_{3,4} = 9.5$ Hz, 1H, H-3), 4.63 (dd, $J_{1,2} = 7.5$ Hz, 1 H, H-1 β), 4.22 (dd, $J_{gem} = 10.2$ Hz, $J_{vic} =$ 5.0 Hz, 1 H, H-6a), 3.74 (dd, $J_{vic} = 10.2$ Hz, 1 H, H-6b), 3.64 (m, 1 H, H-4), 3.53 (m, 1H, H-5), 3.28 (m, 1H, H-5); ¹³C NMR (125 MHz, [D₆]DMSO): $\delta = 153.1$ (C=O urethane), 139.2–118.1 (C-Ar), 100.5 (=CH-Pha), 100.3 (=CH-Phβ), 97.7 (C-1β), 93.3 (C-1α), 79.1 (C-4α), 78.6 (C-4β), 74.3 (C-3β), 73.7 (C-2β), 72.1 (C-3α), 70.9 (C-2α), 68.2 (C-6α), 67.9 (C-6β), 65.5 (C-5β), 62.1 (C-5*a*); FAB-MS (MB): *m*/*z*: calcd for C₂₀H₂₁NO₇: 387.1; found: 388 $[M+H]^+$.

4,6-O-Benzylidene-2-O-chloroacetyl-3-O-(N-phenylcarbamoyl)-D-gluco-

pyranosyltrichloroacetimidate (20): Benzylidene acetal 19 (11.4 g, 29.4 mmol) and pyridine (20 mL) were dissolved in dichloromethane (100 mL) and cooled to 0 °C. Chloroacetic acid anhydride (15 g, 87.7 mmol) was added and the reaction was allowed to proceed at 0°C until the starting material disappeared (TLC: hexane/acetone 1.5:1). The solution was diluted with dichloromethane (200 mL) and extracted with aq K2CO3, aq HCl $(3 \times)$ and aq K₂CO₃. After drying (MgSO₄) the solvent was evaporated. The residue (14.6 g) was dissolved in DMF (21 mL) and hydrazine acetate (2.4 g) was added. After complete reaction (TLC: hexane/acetone 1.5:1) the mixture was diluted with dichloromethane (400 mL) and extracted with water $(2 \times)$ and aq KHCO₃. The organic phase was dried (MgSO₄) and concentrated. Subsequently, the hemiacetal was dissolved in dry dichloromethane (100 mL) and cooled to 0°C. Trichloroacetonitrile (8 mL, 54 mmol) and 1,8-diazabicyclo[5.4.0]undec-7ene (DBU) (775 µL, 7.7 mmol) were added. Upon complete consumption of the starting material (TLC: hexane/acetone 1.5:1) the solution was evaporated to dryness and the residue was purified by flash chromatography (hexane/acetone 2:1) to give the imidate 20 (9.77 g, 54.6%). $R_{\rm f}$ bischloroacetate = 0.44 (hexane/acetone 1.5:1); $R_{\rm f}$ hemiacetal = 0.33 (hexane/ acetone 1.5:1); $R_{\rm f}$ imidate = 0.47 (hexane/acetone 1.5:1); ¹H NMR (500 MHz, [D₆]DMSO): $\delta = 9.97$ (s, 1H, C=NH), 9.78 (s, 1H, NH urethane), 7.49-6.96 (m, 10 H, Ph), 6.53 (d, $J_{12} = 3.6$ Hz, 1 H, H-1 α), 5.71 (s, 1H, =CH-Ph), 5.49(dd, J₂₃ = J₃₄ = 9.8 Hz, 1H, H-3), 5.30 (dd, 1H, H-2), $4.42 - 4.27 (m, 3 H, CH_2Cl, H-6a), 4.11 (dd, J_{4,5} = 9.6 Hz, 1 H, H-4), 4.01 (m, J_{4,5} = 9.6 Hz, 1 H, H-4)$ 1 H, H-5), 3.89 (dd, $J_{gem} = J_{vic} = 10.2$ Hz, H-6b); ¹³C NMR (125 MHz, [D₆]DMSO): δ = 166.3 (C=O), 158.0 (C=N), 152.3 (C=O urethane), 138.7 -118.4 (C-Ar), 100.6 (=CH-Ph), 92.5 (C-1a), 77.2 (C-4), 71.4 (C-2), 68.8 (C-3), 67.2 (C-6), 65.3 (C-5), 40.5 (CH2Cl); FAB-MS (MB): m/z: calcd for C₂₄H₂₂Cl₄N₂O₈: 606.0; found: 607 [M+H]+.

4,6-O-Benzylidene-2-O-chloroacetyl-3-O-(N-phenylcarbamoyl)-\beta-D-glucopyranosylfluoride (21): Imidate **20** (2 g, 21.9 mmol) was dissolved in dry dichloromethane (30 mL) in a polyethylene container and cooled to 0 °C. To the stirred solution was added HF/pyridine complex (420 µL). The reaction was complete after 5 min (TLC: hexane/acetone 2:1). Subsequently, the biphasic reaction was diluted with dichloromethane (130 mL), transferred to a Teflon separaratory funnel and neutralized with cold aq KHCO₃. The organic phase was extracted with aq HCl (3 ×) and aq KHCO₃. After concentration the residue was purified by flash chromatography (hexane/acetone 2:1) to yield the fluoride **21** (1.4 g, 91.4%).

$$\begin{split} & [a]_{D}^{23} = -32.2^{\circ} \ (c=1 \ \text{in dichloromethane}); \ R_{\rm f} = 0.44 \ (\text{hexane/acetone 2:1}); \\ ^{1}\text{H NMR} \ (500 \ \text{MHz}, [D_6] \text{DMSO}): \delta = 9.80 \ (\text{s}, 1 \ \text{H}, \text{NH}), 7.45 - 6.96 \ (\text{m}, 10 \ \text{H}, \text{Ph}), 5.79 \ (\text{dd}, \ J_{1,2} = 6.5, \ J_{1,F} = 53.7 \ \text{Hz}, 1 \ \text{H}, \ \text{H} - 1\beta), 5.68 \ (\text{s}, 1 \ \text{H}, = \text{CH-Ph}), \\ & 5.38 \ (\text{dd}, \ J_{2,3} = J_{3,4} = 8.9 \ \text{Hz}, 1 \ \text{H}, \ \text{H} - 3), 5.09 \ (\text{m}, 1 \ \text{H}, \ \text{H} - 2), 4.51, 4.41 \ (\text{2d}, \ J_{\rm gem} = 15.3 \ \text{Hz}, 1 \ \text{H}, \ \text{CH}_2 \ \text{Cl}), 4.34 \ (\text{dd}, \ J_{\rm gem} = 9.9, \ J_{\rm vic} = 4.5 \ \text{Hz}, 1 \ \text{H}, \ \text{H} - 6a), \\ & 4.03 \ (\text{dd}, \ J_{4,5} = 9.4 \ \text{Hz}, 1 \ \text{H}, \ \text{H} - 4), 3.97 \ (\text{m}, 1 \ \text{H}, \ \text{H} - 5), \ (\text{dd}, \ J_{\rm gem} = 9.9, \ J_{\rm vic} = 10.1 \ \text{Hz}, 1 \ \text{H}, \ \text{H} - 6b); \ ^{13}\text{C} \ \text{NMR} \ (125 \ \text{MHz}, \ [D_6] \ \text{DMSO}): \delta = 152.3 \ (\text{C=O} \ \text{urethane}), 138.6, 136.9 \ (\text{C-}i \ \text{Ph}), 129.4 - 118.5 \ (\text{C-Ar}), 106.3 \ (\text{d}, \ J_{\rm c_{1,F}} = 214.1 \ \text{Hz}, \ \text{C} - 1), 100.5 \ (=C\text{H} - \text{Ph}), 79.9 \ (\text{C} - 4), 73.4 \ (\text{d}, \ J_{\rm c_{2,F}} = 25.4 \ \text{Hz}, \ \text{C} - 2), \\ 70.8 \ (\text{d}, \ J_{\text{c}_{3,F}} = 9.5 \ \text{Hz}, \ \text{C} - 3), 67.2 \ (\text{C-6}), 65.1 \ (\text{C-5}), 40.6 \ (\text{CH}_2 \ \text{Cl}); \ \text{FAB-MS} \ (\text{MB}): m/z: \ \text{calcd for } C_{22}\text{H}_2(\text{CFNO}_7; 665.1; \ \text{found}: 666 \ [M+H]^+. \end{split}$$

$O-(4,6-O-Benzylidene-2-O-chloroacetyl-3-O-(N-phenylcarbamoyl)-\beta-D-glucopyranosyl)-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-phthalimido-\beta-D-glucopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-phthalimido-3-D-glucopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-phthalimido-3-D-glucopyranosyl)-3,6-di-O-benzyl-2-phthalimido-3-D-glucopyranosyl)-3,7-di-O-benzyl-2-phthalimido-3-D-glucopyranosyl)-3,7-di-O-benzyl-2-phthalimido-3-D-glucopyranosyl)-3,7-di-O-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benzyl-2-phthalimido-3-D-benz$

copyranosylazide (22): *a)* Starting from **21**: Disaccharide **13** (85 mg, 86 µmol), fluoride **21** (120 mg, 258 µmol) and ground molecular sieves 4 Å (320 mg) were suspended in dry dichloromethane (3 mL) and stirred for 30 min at 0°C. BF₃•OEt₂ (40 µL, 326 µmol) was added and the reaction was allowed to proceed at ambient temperature for 24 h (TLC: hexane/ acetone 1.5:1). Subsequently, the suspension was filtered over celite, washed with dichloromethane and extracted with aq K₂CO₃. The organic phase was dried (MgSO₄), concentrated and the residue was purified by flash chromatography (hexane/acetone 2:1) to furnish the title trisaccharide **22** (56 mg, 45.5%).

b) Starting from 20: Disaccharide 14 (5.8 g, 5.9 mmol), imidate 20 (6.65 g, 10.9 mmol) and ground molecular sieves 4 Å (18 g) were suspended in dry dichloromethane (50 mL) and stirred for 20 min at 0 °C. Trifluoromethanesulfonic acid trimethylsilylester (600 µL, 3.2 mmol) was added and the reaction was monitored for 3 h (TLC: hexane/acetone 1.5:1). Subsequently, the solids were filtered off over Celite, washed with dichloromethane and the organic phase was extracted with dilute K2CO3. The dried solution (MgSO₄) was concentrated and the residue was purified by flash chromatography (hexane/acetone 2:1) to give trisaccharide 22 (5.26 g, 62.4%). $[\alpha]_{\rm D}^{23} = -14.1^{\circ}$ (c = 0.5 in dichloromethane); $R_{\rm f} = 0.33$ (hexane/ acetone 1.5:1); ¹H NMR (500 MHz, $[D_6]DMSO$): $\delta = 9.71$ (s, 1 H, NH), 7.95-7.65 (m, 8H, Pht), 7.50-7.20 (m, 20H, Ar), 7.00-6.78 (m, 10H, Ar), 5.61 (s, 1H, =CH-Ph), 5.30-5.24 (m, 3H, H-1, H-1', H-3"), 4.95 (m, 2H, H-1", H-2"), 4.83 (d, $J_{\rm gem}$ = 12.4 Hz, 1 H, CH₂O), 4.79 (d, $J_{\rm gem}$ = 11.9 Hz, 1 H, CH₂O), 4.63 (d, J_{gem} = 11.6 Hz, 1 H, CH₂O), 4.58 (d, J_{gem} = 12.2 Hz, 1 H, CH_2O), 4.43-4.37 (m, 4H, CH_2O), 4.32 (m, 2H, CH_2Cl), 4.20 (dd, $J_{2,3}$ = $J_{3,4} = 9.8$ Hz, 1 H, H-3'), 4.15 (dd, $J_{vic} = 4.7$ Hz, $J_{gem} = 9.8$ Hz, 1 H, H-6a"), $4.12 - 4.08 (m, 2H, H-3, H-4), 4.03 (m, 1H, H-2'), 3.98 (dd, J_{45} = 8.4 Hz, 1H, H-2')$ H-4'), 3.84-3.80 (m, 3H, H-2, H-4", H-6a'), 3.68-3.60 (m, 3H, H5, H-6b', H-6b"), 3.54 (m, 1 H, H-5"), 3.49 (dd, $J_{vic} < 1.0, J_{oem} = 10.8$ Hz, 1 H, H-6a), 3.41 (m, 1 H, H-5'), 3.36 (dd, $J_{vic} = 3.1$ Hz, 1 H, H-6b); ¹³C NMR (62.5 MHz, $[D_6]DMSO$: $\delta = 167.1$, 166.1 (C=O), 152.5 (C=O urethane), 138.7, 138.3, 138.0, 137.0 (C-i Ar), 134.8 (C-4/5 Pht), 130.6 (C-1/2 Pht), 129.0-126.1 (C-Ar), 123.4 (C-3/6 Pht), 122.3, 118.6 (C-Ar), 100.4 (=CH-Ph), 99.6 (C-1"), 96.4 (C-1'), 84.8 (C-1), 77.6 (C-4", C-4'), 76.5 (C-3'), 76.2 (C-3), 75.7 (C-5), 75.1 (C-4), 74.5 (C-5'), 74.2 (C-2"), 73.9 (CH₂O), 73.7 (CH₂O), 72.3 (CH2O), 71.7 (C-2", CH2O), 67.6 (C-6, C-6'), 67.4 (C-6"), 65.5 (C-5"), 55.8 (C-2'), 54.6 (C-2), 40.6 (CH₂Cl); ESI-MS: m/z: calcd for C₇₈H₇₁ClN₆O₁₉: 1430.45; found: 1453.50 [M+Na]+.

$O-(4,6-O-Benzylidene-3-O-(N-phenylcarbamoyl)-\beta-D-glucopyranosyl)-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)-$

 $(1 \rightarrow 4)$ -3,6-di-*O*-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosylazide (17): *a*) Starting from 22: Trisaccharide 22 (14 g, 9.78 mmol) was dissolved in methanol/dichloromethane 1:2 (120 mL). To the stirred solution was added K₂CO₃ (300 mg). After complete deacylation (TLC: hexane/acetone 1.5:1) the solids were filtered off, followed by neutralization with acetic acid. The solution was concentrated and purified by flash chromatography (hexane/acetone 1.5:1) furnishing the trisaccharide 17 (11.82 g, 89.2%).

b) Starting from 16: Trisaccharide 16 (3.25 g, 2.3 mmol) was dissolved in dioxane/methanol 1:1 (80 mL) and cooled to 4° C. K₂CO₃ (400 mg) was added and the suspension was stirred for 36 h. Subsequently, the solids were removed by filtration. The solution was neutralized with Amberlyst 15 ion-exchange resin (H⁺) and concentrated. The residue was dissolved in acetonitrile (50 mL) and benzaldehyde dimethylacetal (10 mL) and *p*-toluenesulfonic acid (100 mg) were added. After 1 h the reaction was stopped with triethylamine (150 µL), concentrated and purified by flash chromatography (hexane/acetone 2.5:1) to give the trisaccharide 17

(1.395 g, 44.1%). $[a]_{\rm D}^{23} = -8.2^{\circ}$ (c = 0.5 in dichloromethane); $R_{\rm f} = 0.28$ (hexane/acetone 1.5:1); ¹H NMR (500 MHz, $[D_6]DMSO$): $\delta = 9.6$ (s, 1 H, NH), 7.95-7.65 (m, 8H, Pht), 7.50-7.20 (m, 20H, Ar), 7.00-6.75 (m, 10H, Ar), 5.92 (d, $J_{2,OH} = 6.0$ Hz, 1H, OH-2"), 5.56 (s, 1H, =CH-Ph), 5.28 (d, $J_{1,2} = 9.2$ Hz, 1 H, H-1 β), 5.27 (d, $J_{1,2} = 8.3$ Hz, 1 H, H-1' β), 5.01 (dd, $J_{2,3} = 100$ $J_{3,4} = 9.5$ Hz, 1 H, H-3"), 4.85 (d, $J_{gem} = 12.4$ Hz, 1 H, CH₂O), 4.79 (d, J_{gem} = 12.4 Hz, 1 H, CH₂O), 4.79 (d, J_{gem} = 12.4 Hz, 1 H, CH₂O), 4.79 (d, J_{gem} = 12.4 Hz, 1 H, CH₂O), 4.79 (d, J_{gem} = 12.4 Hz, 1 H, CH₂O), 4.79 (d, J_{gem} = 12.4 Hz, 1 H, 2 Hz, 1 H, 2 Hz, 1 Hz, 11.9 Hz, 1 H, CH₂O), 4.67 (d, $J_{1,2} = 6.9$ Hz, 1 H, H-1" β), 4.65 (d, $J_{gem} =$ 11.6 Hz, 1 H, CH₂O), 4.55 (d, $J_{gem} = 12.2$ Hz, 1 H, CH₂O), 4.42-4.34 (m, 4H, CH₂O), 4.27 (dd, *J*_{2,3} = 10.4, *J*_{3,4} = 8.9 Hz, 1H, H-3'), 4.14-3.96 (m, 6H, H-3, H-4, H-6a', H-6a'', H-2', H-4'), 3.89 (dd, $J_{vic} = 4.2$, $J_{gem} = 11.0$ Hz, 1 H, H-6b'), 3.81 (dd, J_{2,3} = 10.5 Hz, 1 H, H-2), 3.67 - 3.60 (m, 3 H, H-4", H-6b", H-5), 3.54 (m, 1H, H-5′), 3.48 (dd, $J_{\rm vic}$ < 1.0, $J_{\rm gem}$ = 10.6 Hz, 1H, H-6a), 3.45-3.32 (m, 3H, H-2", H-5", H-6b); ¹³C NMR (62.5 MHz, [D₆]DMSO): $\delta = 168.1, 167.0$ (C=O), 152.9 (C=O urethane), 139.1, 138.6, 138.1, 138.0, 137.9, 137.3 (C-i Ar), 134.8 (C-4/5 Pht), 130.6 (C-1/2 Pht), 128.7-126.1 (C-Ar), 123.4 (C-3/6 Pht), 122.3, 118.2 (C-Ar), 103.2 (C-1"), 100.2 (=CH-Ph), 96.5 (C-1'), 84.8 (C-1), 78.1 (C-4"), 77.9 (C-4'), 76.6 (C-3'), 76.2 (C-3), 75.7 (C-5), 75.0 (C-4), 74.7 (C-5'), 74.0 (C-3"), 73.7 (CH₂O), 72.7 (C-2"), 72.1, 71.6 (CH₂O), 67.5 (C-6), 67.5 (C-6', C-6"), 65.7 (C-5"), 56.0 (C-2'), 54.6 (C-2); elemental analysis calcd (%) for C₇₆H₇₀N₆O₁₈ (1355.42): C 67.35, H 5.21, N 6.20; found: C 67.32, H 5.15, N 6.00.

O-(4,6-O-Benzylidene- β -D-mannopyranosyl)-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosylazide (2): Trisaccharide 17 (4.2 g, 3.1 mmol) and pyridine (2 mL) were dissolved in dry dichloromethane (150 mL). The stirred solution was cooled to -40 °C and trifluoromethanesulfonic acid anhydride (700 µL, 4.16 mmol) was added. Upon warming to 0°C the reaction went to completion (TLC: hexane/acetone 1.5:1). The mixture was concentrated in a water bath at ambient temperature followed by evaporation to drynesss in high vacuo. Subsequently, dry DMF (20 mL) and pyridine (2 mL) were added prior to heating to 65 °C. After 2 h the reaction was complete (TLC: dichloromethane/methanol 50:1) and concentrated in high vacuo. The residue was dissolved in dichloromethane (250 mL) and extracted with aq K2CO3. The organic phase was dried (MgSO₄), concentrated to dryness and subsequently dissolved in a mixture of dioxane (20 mL), acetic acid (6 mL) and water (4 mL). After complete hydrolysis of the iminocarbonate (TLC: dichloromethane/methanol 50:1) the solvents were removed at ambient temperature in a rotary evaporator. The residue was taken up in dichloromethane (250 mL), extracted with aq HCl and aq K2CO3, dried (MgSO4) and concentrated. The crude product (carbonate) was dissolved in dry dichloromethane (150 mL) and a freshly prepared solution (5 mL) of sodium (100 mg) in dry methanol (100 mL) was added. After complete reaction (TLC: hexane/acetone 1.5:1) the solution was neutralized with acetic acid, concentrated and purified by flash chromatography (hexane/acetone 1.5:1) to give trisaccharide 2 (2.69 g, 70.2%). $[\alpha]_{D}^{23} = -3.3^{\circ}$ (c = 0.5 in dichloromethane); R_{f} phenylurethane 17 = 0.28 (hexane/acetone 1.5:1); $R_{\rm f}$ triflate = 0.34 (hexane/acetone 1.5:1); $R_{\rm f}$ triflate **17 a** = 0.67 (dichloromethane/methanol 50:1); $R_{\rm f}$ iminocarbonate 17b = 0.62 (dichloromethane/methanol 50:1); $R_{\rm f}$ carbonate 17c = 0.64(dichloromethane/methanol 50:1): $R_{\rm f}$ carbonate 17c = 0.29 (hexane/acetone 1.5:1); $R_{\rm f}$ diol **2**=0.21 (hexane/acetone 1.5:1); ¹H NMR (500 MHz, $[D_6]DMSO$: $\delta = 7.95 - 7.65$ (m, 8H, Pht), 7.41 - 7.22 (m, 15H, Ar), 6.96 -6.75 (m, 10H, Ar), 5.51 (s, 1H, =CH-Ph), 5.28 (d, $J_{1,2}$ = 9.5 Hz, 1H, H-1 β), 5.25 (d, $J_{12} = 8.5$ Hz, 1 H, H-1' β), 4.96 (d, $J_{2.0H} = 4.3$ Hz, 1 H, OH-2"), 4.92 $(d, J_{3,OH} = 6.9 \text{ Hz}, 1 \text{ H}, \text{OH-3''}), 4.82 (d, J_{gem} = 12.1 \text{ Hz}, 2 \text{ H}, \text{CH}_2\text{O}), 4.62 (d, M_2)$ $J_{1,2} < 1.0$ Hz, 1 H, H-1" β), 4.60 (d, $J_{gem} = 12.2$ Hz, 1 H, CH₂O), 4.55 (d, $J_{\text{gem}} = 12.2 \text{ Hz}, 1 \text{ H}, \text{CH}_2\text{O}), 4.43 - 4.37 \text{ (m, 4 H, CH}_2\text{O}), 4.21 \text{ (dd, } J_{2.3} = 10.4,$ $J_{3,4} = 8.8 \text{ Hz}, 1 \text{ H}, \text{ H-3'}), 4.14 - 3.96 \text{ (m, 5 H, H-3, H-4, H-6a'', H-2', H-4')},$ 3.80-3.76 (m, 3H, H-2, H-2", H-6a'), 3.71 (dd, $J_{3,4}=J_{4,5}=9.5$ Hz, 1H, H-4"), 3.66-3.47 (m, 5H, H-5, H-6a, H-6b', H-3", H-6b"), 3.41 (m, 1H, H-5'), 3.36 (m,1H, H-6b), 3.10 (m, 1H, H-5"); ¹³C NMR (62.5 MHz, $[D_{6}]DMSO$): $\delta = 168.0, 167.1, 167.0$ (C=O), 138.4, 138.3, 138.1, 137.9 (C-*i* Ar), 134.8 (C-4/5 Pht), 130.7, 130.5 (C-1/2 Pht), 128.7-126.3 (C-Ar), 123.4 (C-3/6 Pht), 100.9 (=CH-Ph), 100.4 (C-1" β , J_{C-1H-1} = 158.6 Hz from a coupled HMQC spectrum), 96.5 (C-1' β , $J_{C-1,H-1} = 169.0$ Hz see above), 84.8 $(C-1\beta, J_{C-1,H-1} = 166.9 \text{ Hz see above}), 78.3 (C-4''), 77.3 (C-4'), 76.2 (C-3), 76.0$ (C-3'), 75.6 (C-5), 74.9 (C-4), 74.5 (C-5'), 73.7, 73.6, 72.1, 71.5 (CH₂O), 70.9 (C-2"),69.9 (C-3"), 67.8 (C-6', C-6"), 67.5 (C-6), 66.8 (C-5"), 56.0 (C-2'), 54.7 (C-2); elemental analysis calcd (%) for C69H65N5O17 (1236.30): C 67.04, H 5.30, N 5.66; found: C 66.70, H 5.18, N 5.52. ESI-MS: m/z: calcd for $C_{69}H_{65}N_5O_{17}$: 1235.44); found: 1258.45 [*M*+Na]⁺.

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